

Small RNA-mediated DNA methylation during plant reproduction

Hiu Tung Chow  and Rebecca A. Mosher *

The School of Plant Sciences, The University of Arizona, Tucson, Arizona 85721-0036, USA

*Author for correspondence: rmosher@arizona.edu

Both authors contributed to writing the article.

The author responsible for the distribution of materials integral to the findings presented in this article in accordance with the policy described in the Instructions for Authors (<https://academic.oup.com/plcell>) is: Rebecca A. Mosher (rmosher@arizona.edu).

Review

Abstract

Reproductive tissues are a rich source of small RNAs, including several classes of short interfering (si)RNAs that are restricted to this stage of development. In addition to RNA polymerase IV-dependent 24-nt siRNAs that trigger canonical RNA-directed DNA methylation, abundant reproductive-specific siRNAs are produced from companion cells adjacent to the developing germ line or zygote and may move intercellularly before inducing methylation. In some cases, these siRNAs are produced via non-canonical biosynthesis mechanisms or from sequences with little similarity to transposons. While the precise role of these siRNAs and the methylation they trigger is unclear, they have been implicated in specifying a single megaspore mother cell, silencing transposons in the male germ line, mediating parental dosage conflict to ensure proper endosperm development, hypermethylation of mature embryos, and trans-chromosomal methylation in hybrids. In this review, we summarize the current knowledge of reproductive siRNAs, including their biosynthesis, transport, and function.

Introduction

The most abundant small RNAs in most plant genomes are 24-nt short interfering (si)RNAs that are key components in RNA-directed DNA methylation (RdDM) (Matzke and Mosher, 2014). RdDM begins when RNA polymerase IV (Pol IV) is recruited to DNA through the action of a CLASSY (CLSY) protein (Zhou et al., 2018). Pol IV transcribes a short (25–40 nt) non-coding RNA before backtracking along the DNA, which causes the 3' end of the transcript to be taken up by RNA-DEPENDENT RNA POLYMERASE 2 (RDR2) (Blevins et al., 2015; Zhai et al., 2015a; Fukudome et al., 2021). Using the Pol IV transcript as a template, RDR2 transcribes a complementary strand, producing double-stranded RNA (Singh et al., 2019; Huang et al., 2021). These short double-stranded RNAs are trimmed to 24-nt siRNA duplexes by DICER-LIKE 3 (DCL3) (Wang et al., 2021; Loffer et al., 2022). One strand of the siRNA duplex is bound by an ARGONAUTE (AGO) and directs these effector proteins to complementary transcripts produced

by RNA polymerase V (Pol V), or possibly to single-stranded DNA resulting from Pol V transcription (Wierzbicki et al., 2008, 2009; Lahmy et al., 2016; Liu et al., 2018). The siRNA/AGO complex then recruits DOMAINS REARRANGED METHYLTRANSFERASE (DRM) to catalyze cytosine methylation in all sequence contexts (Zemach et al., 2013; Stroud et al., 2014; Zhong et al., 2014). This DNA methylation helps recruit Pol IV for additional siRNA production (Law et al., 2013; Choi et al., 2021), thereby creating a stable feedback loop. Because non-symmetric DNA methylation is lost from one daughter strand during semi-conservative DNA methylation, RdDM's feedback loop is important for maintaining CHH methylation (where H=A, T, or C) in euchromatin (Law and Jacobsen, 2010; Law et al., 2013). In addition to this canonical mechanism, variations of RdDM initiate methylation at unmethylated sequences (Nuthikattu et al., 2013; McCue et al., 2015; Cuerda-Gil and Slotkin, 2016; Sigman et al., 2021).

RdDM primarily functions at transposons, including both DNA- and RNA-based elements (Kasschau et al., 2007; Zhang et al., 2007; Mosher et al., 2008), and is responsible for transcriptional silencing of transposons at the boundary between heterochromatin and euchromatin (Li et al., 2015; Böhmendorfer et al., 2016). RdDM of transposons that are in proximity to protein-coding genes can influence expression of such genes (Hollister and Gaut, 2009) and remnants of transposons can create gene-specific regulatory structures that silence gene expression (Chan et al., 2006; Kinoshita et al., 2007; Henderson and Jacobsen, 2008). Differing activity of RdDM in alternate developmental contexts thus provides epigenetic control of gene expression (Vu et al., 2013).

In angiosperms, 24-nt siRNAs are most abundant in reproductive tissues (Mosher et al., 2009; Grover et al., 2020; Zhou et al., 2022a), which are a complex mix of somatic, germ line, and zygotic cells. The female and male germ lines are established when megaspore or microspore mother cells, respectively, are specified from surrounding somatic cells (Berger and Twell, 2011). The megaspore and microspore mother cells undergo meiosis to generate the haploid megaspore and microspore, which then undergo mitosis to generate female and male gametophytes, respectively. In most angiosperms, the megaspore goes through three rounds of nuclear division before cytokinesis to generate the 7-celled female gametophyte, while the microspore has an asymmetric division, followed by a second division of the smaller cell to create the male gametophyte (pollen grain). The two haploid sperm cells in the pollen grain fertilize the haploid egg cell and diploid central cell to create the zygotes that develop into the embryo and endosperm, respectively. Mega/microspore specification, meiosis, gametogenesis, fertilization, and zygotic development all happen in intimate connection with diploid somatic cells, providing the potential for intercellular, and indeed intergenerational, movement of siRNAs during reproduction (Feng et al., 2013).

Consistent with the abundance of 24-nt siRNAs in reproductive tissues, RdDM is required for reproductive development in a variety of species, including tomato (Gouil and Baulcombe, 2016), other *Brassicaceae* (Grover et al., 2018; Wang et al., 2020), rice (Xu et al., 2020; Zheng et al., 2021; Chakraborty et al., 2022; Wang et al., 2022), and maize (Erhard et al., 2009). Here, we will discuss the role of RdDM in establishing and maintaining epigenetic marks during plant reproduction, with particular emphasis on the role of small RNA movement from somatic cells.

Male germ line

Although there is no defect in *Arabidopsis thaliana* when RdDM is eliminated, research in multiple other species suggests that 24-nt or transposon-associated siRNAs are required for male germ line development (Walker et al., 2018; Bélanger et al., 2020; Teng et al., 2020; Wang et al., 2020; Nan et al., 2022). During development, microspore mother cells, microspores, and immature pollen

are surrounded by a layer of sporophytic cells called the tapetum or tapetal nurse cells (Figure 1A). Likewise, after pollen development, sperm cells are encased within the vegetative cell. While they do not contribute *genetically* to the next generation, these companion cells frequently produce male-reproductive specific siRNAs that move intercellularly to trigger methylation or gene silencing in the germ line, causing an *epigenetic* impact on the subsequent generation.

Nurse cell siRNAs

During *Arabidopsis* male germ line development, abundant 24-nt siRNAs accumulate from several hundred loci (Figure 1B) (Long et al., 2021). These siRNAs require *RDR2* and *CLSY3*, the latter being expressed in the tapetal nurse cells surrounding the microspore mother cells (Long et al., 2021). Tapetal-specific expression of *RDR2* in the *rdr2* background is sufficient for siRNA accumulation in the microspore mother cells, indicating that these “nurse cell siRNAs” are produced specifically in the tapetum before moving intercellularly into microspore mother cells (Long et al., 2021). As expected, the nurse cell siRNAs induce DNA methylation in *cis*, but they also trigger methylation in *trans* at sites with up to three mismatches (Long et al., 2021). This results in CHH hypermethylation of hundreds of loci in microspore mother cells relative to somatic tissues (Walker et al., 2018; Long et al., 2021). These sites remain hypermethylated in microspores, sperm cells, and vegetative cells, likely through an absence of demethylation as well as continued transport of nurse cell siRNAs throughout germ line development (Walker et al., 2018; Long et al., 2021).

Unlike somatic RdDM, which mostly occurs at transposons and transposon fragments, most male germ line-specific hypermethylated loci overlap genes, suggesting that nurse cell siRNAs regulate gene expression during germ line development (Walker et al., 2018). In support of this hypothesis, some of the hypermethylated genes are transcriptionally up-regulated in *drm* microspore mother cells but not in *drm* leaves (Walker et al., 2018). In addition to transcriptional silencing of targets, *drm* mutation also causes mis-splicing of *MPS1*, a gene targeted by RdDM in the male germ line. This mis-splicing results in defective meiosis (Walker et al., 2018), highlighting the functional significance of nurse cell siRNA-mediated RdDM in the male germ line. However, it is not clear whether nurse cell *trans*-methylation is developmentally significant for most target genes.

Reproductive phasiRNAs

While nurse cell siRNA loci have been described only in *Arabidopsis* to date, a different class of highly expressed siRNA loci are present during male germ line development in many other angiosperms—phased, secondary, small interfering RNAs (phasiRNAs) (Figure 1C) (Johnson et al., 2009; Song et al., 2012). Biosynthesis of phasiRNAs is distinct from Pol IV-dependent siRNAs. First, Pol II transcribes *PHAS* loci and generates long precursor transcripts, which are targeted by microRNAs for cleavage. The cleaved

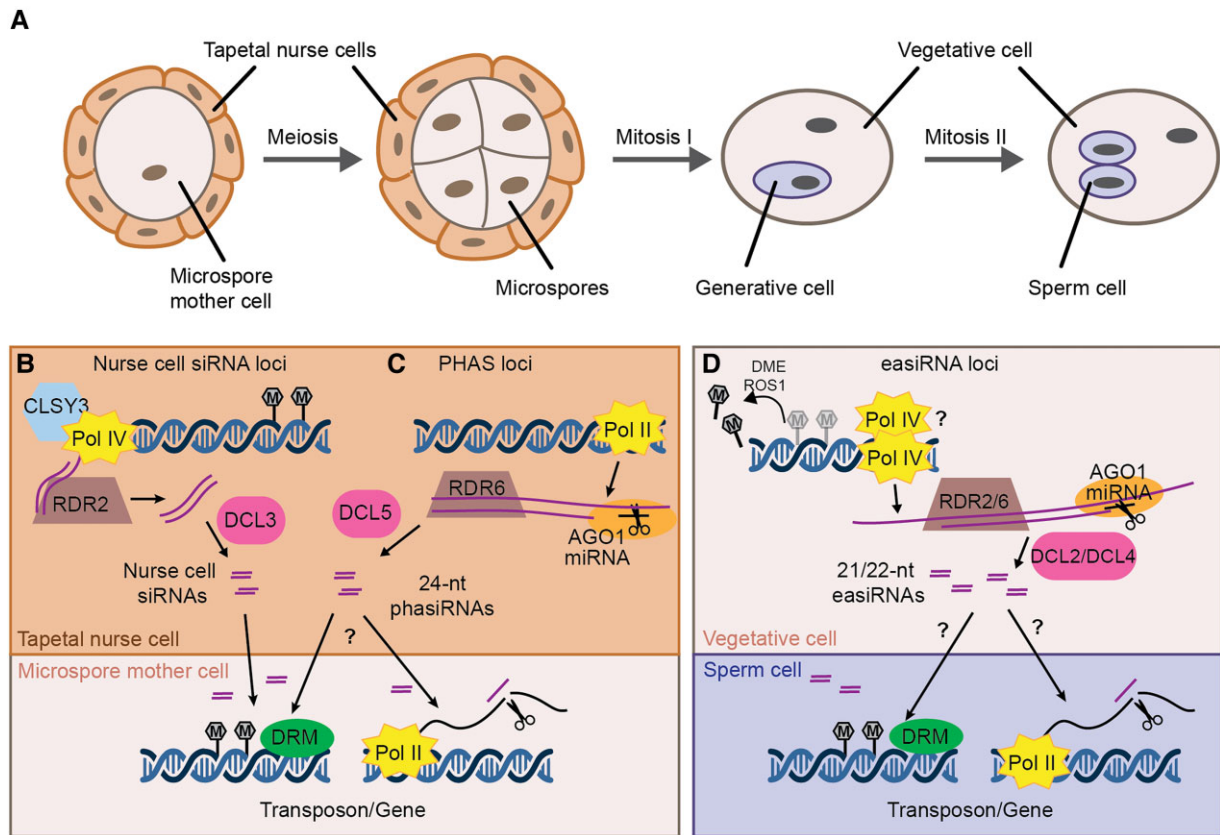


Figure 1 Male reproductive development, siRNA biogenesis, and proposed intercellular movement. **A**, Male germ line development. A microspore mother cell undergoes meiosis to generate four haploid microspores (shown in tetrad stage). Each microspore has an asymmetric mitotic division to create the vegetative cell and the generative cell. The generative cell later divides to create the two sperm cells. Together, the vegetative cell and sperm cells form the male gametophyte or mature pollen grain. Microspore mother cells, microspores, and immature pollen are surrounded by a layer of sporophytic cells called the tapetum or tapetal nurse cells. **B**, The Arabidopsis tapetum produces 24-nt nurse cell siRNAs through the canonical RdDM pathway. Pol IV and CLSY3 produce RNA precursors which are converted into dsRNA by RDR2 before processing by DCL3 to generate 24-nt nurse cell siRNAs. Nurse-cell siRNAs move into microspore mother cells, inducing DNA methylation with the aid of DRM1 and/or DRM2 (DRM). **C**, In the tapetum of many other angiosperms, reproductive phasiRNAs are produced from *PHAS* loci. Pol II transcribes *PHAS* loci to generate precursor transcripts, which are targeted by miRNAs for cleavage. The cleaved transcripts are converted into dsRNA by RDR6, and those targeted by miR2275 are further processed by DCL5 to generate 24-nt phasiRNAs. These phasiRNAs move intercellularly into germ cells, and might induce DNA methylation (left arrow) or post-transcriptional gene silencing through transcript cleavage (right arrow). **D**, Epigenetically-activated (ea)siRNAs are produced from reactivated transposons in the vegetative nucleus, where DME and ROS1 actively demethylate transposons and some protein coding genes. The demethylated loci are transcribed by either Pol II or Pol IV and the resulting RNAs are cleaved by miRNA/AGO complexes, triggering their conversion into dsRNA by RDR2 or/and RDR6. DCL2 and DCL4 then produce 22- and 21-nt easiRNAs from the dsRNA, respectively. These vegetative cell-derived easiRNAs can induce gene silencing in the sperm cell, either through transcriptional gene silencing (left arrow) or post-transcriptional gene silencing (right arrow).

transcripts are converted into dsRNA by RDR6, and further processed by DCLs to generate phasiRNAs. There are two major classes of reproductive phasiRNAs: 21-nt phasiRNAs are usually produced by DCL4 following miR2118 cleavage, while miR2275 and DCL5 are responsible for generating most 24-nt phasiRNAs (Song et al., 2012; Zhai et al., 2015b; Pokhrel et al., 2021). Originally identified in rice and other monocots, reproductive phasiRNAs are also present in many eudicot lineages, although they have been lost from some families, including the well-studied Brassicaceae and Fabaceae (Xia et al., 2019). Their presence in *Amborella trichopoda* indicates conservation of this pathway since the

emergence of angiosperms, however the number and sequence of *PHAS* loci is variable across species and some lineages use different microRNA triggers, or other strategies to induce dsRNA production (Kakrana et al., 2018; Xia et al., 2019; Bélanger et al., 2020; Pokhrel and Meyers, 2022). This conservation of phasiRNA production but variation in phasiRNA sequences is reminiscent of Pol IV-dependent siRNAs.

Both 21- and 24-nt reproductive phasiRNAs display distinct spatiotemporal distribution throughout male germ line development. While 21-nt phasiRNAs are abundant before meiosis and localize in all anther cell layers, most 24-nt

phasiRNAs express during meiosis and persist throughout post-meiotic development (Zhai et al., 2015b; Araki et al., 2020). In maize, 21-nt phasiRNAs are absent from mutants that lack an epidermis, suggesting that they are synthesized in these cells, while the tapetum, but not microspore mother cells, is required for producing 24-nt phasiRNAs (Zhai et al., 2015b). While 24-nt phasiRNAs are detected in tapetum and other somatic cells by fluorescent in situ hybridization, their levels are highest in microspore mother cells, indicating that these tapetal-derived siRNAs move intercellularly into germ cells (Zhou et al., 2022b). Whether 24-nt reproductive phasiRNAs have the same function in most species as nurse cell siRNAs have in Brassicaceae remains to be explored.

While 21-nt pre-meiotic phasiRNAs cause post-transcriptional regulation, 24-nt meiotic phasiRNAs are proposed to induce DNA methylation (Jiang et al., 2020; Zhang et al., 2021). In maize meiotic anthers, 24-nt *PHAS* loci are highly methylated at CHH contexts and that methylation is depleted in *dcl5* mutant anthers, indicating that 24-nt phasiRNAs induce methylation in cis (Zhang et al., 2021). Whether 24-nt phasiRNAs can also trigger methylation in trans to impact gene expression is less clear. In rice, some 24-nt phasiRNAs match to the promoter of *CK11*, a casein kinase, and this promoter has panicle-specific CHH methylation (Yu et al., 2021). However, it is unknown if this methylation is directed by 24-nt phasiRNAs or Pol IV-dependent siRNAs produced from the methylated *PHAS* locus in parallel. Investigating if 24-nt meiotic phasiRNAs target genes in trans and the rules for this targeting will be an important area for future research.

Although their mechanism of action is unknown, it is clear that 24-nt phasiRNAs are central to male fertility during stress conditions. Maize *dcl5*, which is defective in 24-nt phasiRNAs production, exhibits short anthers with defective tapetal cells when grown at higher temperatures, suggesting that 24-nt phasiRNAs might play a role in conferring male fertility during heat stress (Teng et al., 2020). Loss of the transcription factors necessary for *PHAS* and *DCL5* expression also causes male sterility in maize (Nan et al., 2022). Loss of 21-nt reproductive phasiRNAs also causes environment-dependent sterility, indicating that both size classes of reproductive phasiRNAs influence fertility in the face of abiotic stress (Zhai et al., 2015b; Araki et al., 2020; Yadava et al., 2021). Because many transposons respond to environmental signals, it might be that reproductive phasiRNAs are required to transcriptionally or post-transcriptionally silence such transposons, and therefore phenotypes due to loss of reproductive phasiRNAs are only visible in conditions when the transposons are active.

easiRNAs

While 24-nt nurse cell siRNAs and 24-nt phasiRNAs are produced from the tapetum during meiosis and may regulate protein-coding genes, a different class of siRNA is produced from the male gametophyte and proposed to control transposons. Epigenetically activated small interfering RNAs

(easiRNAs) are 21/22-nt siRNAs that are generated from reactivated transposons in the vegetative cell (Figure 1D; Slotkin et al., 2009; Martínez et al., 2016). In Arabidopsis, the vegetative cell is actively demethylated by demeter (DME) and repressor of silencing 1 (ROS1), related DNA glycosylases that demethylate transposons and some protein-coding genes in the vegetative nucleus (Schoft et al., 2011; Ibarra et al., 2012; Park et al., 2017; Khouider et al., 2021). At the same time, decrease in DNA methylation 1 (DDM1) is lower in the vegetative cell than the sperm cells, further relaxing heterochromatin (Slotkin et al., 2009). Homologous glycosylases demethylate the vegetative cell in rice, indicating that this process is evolutionarily conserved (Kim et al., 2019). Demethylation and relaxation of heterochromatin makes transposons more accessible, and either Pol II or Pol IV transcribe easiRNA precursors in the vegetative nucleus (Slotkin et al., 2009; Borges et al., 2018). Like *PHAS* transcripts, these precursors are recognized by 21- or 22-nt miRNAs, which mediate cleavage with the aid of AGO1 (Creasey et al., 2014; Borges et al., 2018). The cleaved transposon transcripts are then converted into double-stranded RNA (dsRNA) by an RDR, although it is unclear whether RDR2 or RDR6 are involved in this process (Creasey et al., 2014; Martínez et al., 2018; Wang et al., 2020). The resulting dsRNAs are processed by DCL4 and DCL2, producing 21- and 22-nt easiRNAs, respectively.

There is evidence that vegetative cell-derived easiRNAs move intercellularly and function within sperm cells. Purified sperm cells contain abundant 21-nt retrotransposon siRNAs but lack the precursor transposon transcripts, suggesting that easiRNAs are not produced in sperm cells (Slotkin et al., 2009). Similarly, loss of *DME* in vegetative cells is associated with reduced CHH methylation in sperm cells (Ibarra et al., 2012), suggesting that siRNAs that are produced in vegetative cells act in sperm cells. Movement of siRNAs between the vegetative cell and sperm cells was directly demonstrated by producing siRNAs in the vegetative cell from a truncated green fluorescent protein (GFP) sequence. These siRNAs could silence a full-length GFP reporter that was specifically expressed in sperm cells, demonstrating their non-cell autonomous function (Martínez et al., 2016). Similarly, the sperm cell-specific GFP reporter was silenced when its 3' untranslated region contained binding sites for endogenous easiRNAs, and this silencing was eliminated when the 2b protein, which sequesters siRNAs, was expressed in vegetative cells (Martínez et al., 2016). Together, these experiments demonstrate that easiRNAs produced in the vegetative cell can move into sperm cells and silence gene expression there. Although it is unclear whether easiRNAs trigger DNA methylation or post-transcriptional gene silencing, they are proposed to silence transposons to prevent new insertions that would be passed to the next generation (Slotkin et al., 2009). EasiRNAs are also implicated in parental dosage balance in the endosperm after fertilization (discussed further below) (Borges et al., 2018; Martínez et al., 2018).

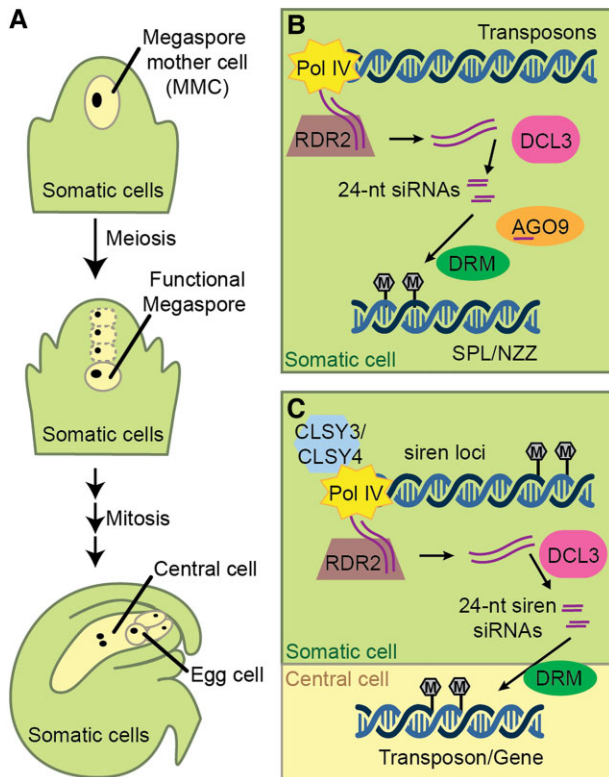


Figure 2 Female reproductive development and the action of maternal 24-nt reproductive siRNAs. **A**, Female germ line development. Within each ovule, a single MMC undergoes meiosis to generate four haploid megaspores. Three of these degenerate, leaving a single functional megaspore. The megaspore goes through three rounds of nuclear division before cytokinesis to generate a 7-celled female gametophyte (antipodal cells not shown). The female gametophyte contains a binucleate central cell and a haploid egg cell that are ready for fertilization. The female germ line is surrounded by somatic cells throughout this development. **B**, Before meiosis, Pol IV, RDR2, and DCL3 produce 24-nt siRNAs, which interact with AGO9 to mediate DNA methylation and transcriptional gene silencing of SPL/NZZ via DRM1 and/or DRM2 (DRM). Regulation of SPL/NZZ is required for the specification of a single MMC. **C**, In somatic cells of a mature ovule, CLSY3 and CLSY4 direct Pol IV to transcribe siren loci. These transcripts are converted into double stranded by RDR2 and further processed into 24-nt siren siRNAs by DCL3. siren siRNAs induce methylation at protein coding genes in somatic cells and are proposed to move intercellularly, causing methylation in the gametophyte.

Female germ line

Because they are buried in maternal somatic tissue (Figure 2A), it is more difficult to study siRNAs and the impact of RdDM on female germ line development. Genetic evidence demonstrates that proteins associated with RdDM are involved in specifying a single megaspore mother cell (MMC) and thereby initiating the female germ line (Figure 2B) (Olmedo-Monfil et al., 2010; Mendes et al., 2020). In Arabidopsis, loss of *RDR2*, *DCL3*, *AGO9*, or *DRM* results in multiple MMC-like cells per ovule (Olmedo-Monfil et al., 2010; Mendes et al., 2020). Because *AGO9* accumulates

specifically in somatic cells surrounding the MMC, these phenotypes suggest that RdDM acts in somatic cells to restrict female germ line identity. Multiple MMC-like cells arise due to ectopic expression of sporocyteless/nozzle (*SPL/NZZ*), which has an increased expression domain in *ago9* and *drm1 drm2* mutants (Mendes et al., 2020). The requirement for *DRM* function suggests that *SPL/NZZ* regulation is through transcriptional gene silencing. Trans-acting siRNAs, which cause post-transcriptional gene silencing, are also required to limit female germ line initiation (Su et al., 2017, 2020), suggesting that multiple small RNA pathways interact at this critical point of development.

Consistent with RdDM's proposed role in maintaining somatic cell identity (i.e. repressing germ line identity), *AGO9* accumulates in somatic cells surrounding the haploid megaspore and the developing female gametophyte. However, *AGO9* is also required to silence transposons in the developing ovules (Olmedo-Monfil et al., 2010). This observation is similar to intercellular movement of nurse cell siRNAs from the tapetum to the microspores suggesting siRNA movement might occur from the soma to germ line during both male and female reproductive development.

Like the tapetum, somatic tissue surrounding the female germ line expresses abundant 24-nt siRNAs from a modest number of loci (Rodrigues et al., 2013; Grover et al., 2020; Zhou et al., 2022a). Known as siren loci due to their original description as siRNAs in endosperm, these siRNAs are produced by RNA Pol IV in ovules before fertilization and from the maternal somatic seed coat after fertilization (Figure 2C) (Rodrigues et al., 2013; Grover et al., 2020). Loci expressing siren siRNAs are present in both monocots and dicots, but there is little conservation of individual loci, even between species within the same family (Grover et al., 2020).

Like nurse cell siRNAs, Pol IV transcription of siren loci is *CLSY3*- and *CLSY4*-dependent, with the former having a dominant role (Zhou et al., 2022a). However, there is only a small overlap between loci producing these siRNAs (12 shared loci among 797 nurse cell and 68 siren loci), suggesting that control of siRNA expression in somatic cells surrounding the male and female germ lines is distinct in some way. While they are largely produced from different loci, siren siRNAs also induce methylation in trans at protein coding genes and cause altered gene expression at a subset of target genes (Burgess et al., 2022). However, it is not yet known whether this methylation occurs in the germ line or is restricted to maternal somatic cells. Live cell imaging of methylation in Arabidopsis revealed that global CHH methylation in the egg cell requires Pol V but not Pol IV, indicating that siRNAs might be made elsewhere and transported into the egg cell to induce methylation (Ingouff et al., 2017). Consistent with this hypothesis, rice egg cells and ovaries have highly similar siRNA patterns. Both cell types accumulate abundant siRNAs from a small number of discreet loci, suggesting that siren siRNAs might move from the maternal soma to the gametophyte in rice (Li et al., 2020). While these observations hint that siren siRNAs might be transported

from the maternal soma to the germ line, direct evidence in support of this hypothesis is lacking.

Seed development

Double fertilization in angiosperms results in two fertilization products, the embryo and the endosperm, that carry the same genetic information but differ in ploidy (Figure 3A). The embryo grows into the next generation and therefore maintaining epigenetic control of transposons is critical in this tissue. In contrast, transposon control is less important in the ephemeral endosperm, which will ultimately be consumed by the embryo. However, recent research indicates that RdDM also plays a role during endosperm development by regulating expression of dosage-sensitive growth factors (Lu et al., 2012; Kirkbride et al., 2019).

Endosperm: balancing parental contributions

Because the endosperm forms from fertilization of the diploid central cell by a haploid sperm cell, it is triploid with a 2:1 matrigenic:patrigenic ratio. (We use “matrigenic” and “patrigenic” to distinguish the maternally or paternally transmitted alleles in the zygote from the true maternal and paternal alleles in the sporophyte and gametophytes (Queller and Strassmann, 2002).) Maintenance of this 2 matrigenic:1 patrigenic dosage is

important for successful endosperm and seed development, and an imbalance in parental dosage is a key mechanism of reproductive isolation following polyploidization (Müntzing, 1933; Scott et al., 1998). The Parental Conflict Hypothesis describes the fact that matrigenic alleles benefit by sharing resources with their maternal half-siblings, while patrigenic alleles benefit by extracting maximum resources from the maternal sporophyte (Haig and Westoby, 1989). This conflict might be resolved by the evolution of genomic imprinting, where the expression of a gene depends on its parent of origin. Parental Conflict Hypothesis therefore predicts that matrigenically expressed genes (MEGs) tend to restrict growth while patrigenically expressed genes (PEGs) generally enhance growth, and balance between MEGs and PEGs is necessary for successful endosperm development. Expression or activity of imprinted genes might vary in different species (independent of polyploidization), creating an imbalance in *effective* parental dosage (also known as the Endosperm Balance Number) (Johnston et al., 1980).

There is accumulating evidence that RdDM influences endosperm development by mediating expression differences between the matrigenic and patrigenic genomes. Compared to the embryo, endosperm is enriched for siRNAs mapping to genes (Erdmann et al., 2017) and thousands of genes are misexpressed in endosperm lacking Pol IV (Satyaki and Gehring, 2022). Imprinted genes are

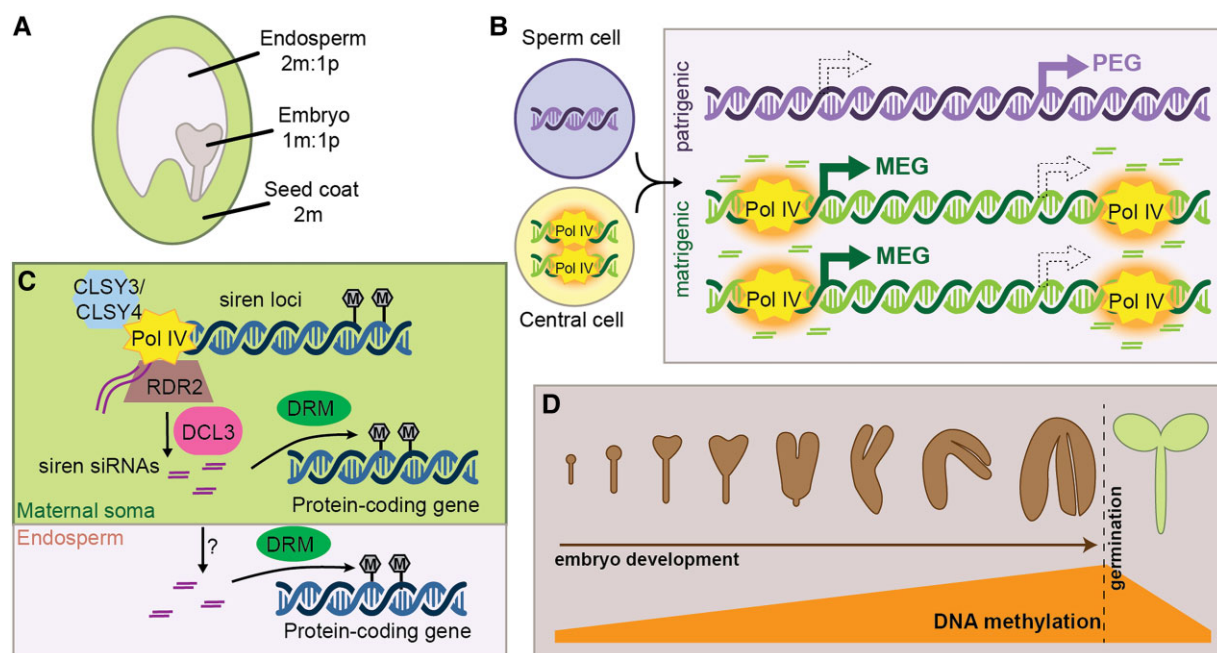


Figure 3 24-nt siRNA production during seed development. A, The developing seed is composed of three tissues, with distinct genetic complements. m, maternal or matrigenic; p, patrigenic. B, Pol IV activity in the central cell is hypothesized to establish an epigenetic state that is maintained on matrigenic chromosomes in the endosperm (orange halo). This epigenetic state causes allele-specific siRNA production in the endosperm and might also influence imprinted gene expression. However, whether methylation caused by allele-specific siRNAs reinforces allele-specific gene expression is unclear. (MEG, PEG; filled arrows depict the expressed state while hollow arrows with dashed outline show the non-expressed allele). C, Siren siRNAs are produced in the immature seed coat and trigger DNA methylation at protein-coding genes via DRM proteins. Siren siRNAs might also move intercellularly, resulting in maternally specific accumulation of siRNAs in the endosperm. Siren siRNA methylation of protein-coding genes in the endosperm might influence seed development. D, During embryo development, canonical RdDM is upregulated at many transposons, resulting in hypermethylation of the genome in mature embryos. This methylation is rapidly lost upon germination.

associated with parentally biased siRNA loci (Rodrigues et al., 2013; Xin et al., 2014; Pignatta et al., 2015; Erdmann et al., 2017), leading to the hypothesis that allele-specific siRNAs help repress transcription at the non-expressed allele (Figure 3B). However, it is not clear how siRNAs might differentially act on the homologous alleles. Additionally, in maize, siRNAs are frequently associated with the expressed allele (Xin et al., 2014), suggesting that allele-specific expression of siRNAs and genes might be driven in parallel by a single parent-specific epigenetic state. There is evidence that RdDM is required for establishing such an epigenetic state before fertilization, particularly in the female germ line. Over 4,500 genes are misexpressed in heterozygous endosperm produced from Pol IV mutant mothers, and these genes are enriched for MEGs and PEGs (Satyaki and Gehring, 2022). Similarly, maternally biased siRNA regions are overwhelmingly downregulated in endosperm from Pol IV mutant mothers (Satyaki and Gehring, 2022). In one example, loss of Pol IV in mothers results in loss of maternal siRNAs at the *Agamous*-like transcription factor *AGL91* (a PEG) and derepression of the maternally derived *AGL91* allele (Kirkbride et al., 2019). Together, these observations suggest that RdDM might establish a heritable epigenetic state before fertilization that persists and impacts siRNA and gene expression after fertilization. However, identification of such epigenetic states has remained elusive (Satyaki and Gehring, 2022). Alternatively, rather than establishing a heritable epigenetic state (or in addition to such establishment), siRNAs might be produced in the maternal somatic tissue, and transported into endosperm where they accumulate and regulate gene expression (Grover et al., 2020, discussed further below).

Further support for the hypothesis that RdDM establishes epigenetic states in gametes comes from the observation that loss of RdDM components such as Pol IV in tetraploid pollen donors allows successful interploidy hybridization with diploid mothers (Erdmann et al., 2017; Martinez et al., 2018; Satyaki and Gehring, 2019), while loss of maternal Pol IV exacerbates seed lethality in such paternal excess crosses (Satyaki and Gehring, 2022). These observations suggest that Pol IV-dependent siRNAs, or the methylation marks they establish, increase the effective dosage of both maternally and paternally derived genomes. The nature of effective dosage is unknown, but might be related to expression of MEGs and PEGs (Brandvain and Haig, 2018; Raunsgard et al., 2018). Consistent with this hypothesis, disruption of several PEGs restores viability to endosperm with excess paternally derived genomes (Kradolfer et al., 2013; Wolff et al., 2015) and many PEGs are upregulated in paternal-excess endosperm beyond the two-fold that is expected due to increased copy number (Martinez et al., 2018; Satyaki and Gehring, 2019). Upregulation of these PEGs might be due to loss of CHH methylation at nearby transposons in paternal excess endosperm, a process that unexpectedly requires Pol IV activity in pollen (Martinez et al., 2018). This counterintuitive observation is reconciled by hypothesizing that Pol IV-transcripts are

cleaved by alternative DCL proteins to produce 21/22-nt easiRNAs in the male germ line, and that these shorter siRNAs compete with Pol IV-dependent 24-nt siRNAs (Borges et al., 2018; Martinez et al., 2018; Panda et al., 2020; Wang et al., 2020). Under this model, the additional genomes in tetraploid fathers produce an excess of 21/22-nt siRNAs, which overwhelm maternally or maternally derived 24-nt siRNAs, causing demethylation, misexpression of PEGs, and endosperm failure (Borges et al., 2018; Martinez et al., 2018). However, whether 21/22-nt siRNAs produced in pollen are transmitted to the zygote where they might encounter maternally derived 24-nt siRNAs is unclear (discussed in “Outstanding Questions”).

In addition, there is a substantial debate regarding the nature of 24-nt siRNAs in endosperm, with various publications reporting either an extensive maternally derived bias (Mosher et al., 2009; Grover et al., 2020) or limited parental bias (Rodrigues et al., 2013; Xin et al., 2014; Erdmann et al., 2017; Satyaki and Gehring, 2022). The different conclusions are correlated with differing approaches for measuring siRNAs: assessing the many thousands of individual siRNA loci identifies only limited examples of either maternally or paternally derived bias, while considering the siRNA population as a whole yields substantial maternally derived bias. This discrepancy is likely due to accumulation of maternally derived siRNAs in endosperm (Grover et al., 2020). Although they are produced from only 1% to 2% of all loci, maternally derived siRNAs are highly expressed and form a substantial fraction of the total siRNA population. Given their abundant expression in maternal somatic tissue, the presence of maternally derived siRNAs in endosperm samples could result from contamination of endosperm with maternal tissue before sequencing (Schon and Nodine, 2017). However, these maternally derived siRNAs are also found in laser microdissected samples, which are unlikely to contain maternal contamination (Grover et al., 2020). This observation suggests that maternally derived expression (or transport from the maternal soma, as discussed below) is limited in the number of loci but extensive in the impact on siRNA transcriptome.

Intercellular RdDM in seed development

While siRNAs from most loci accumulate normally in Pol IV heterozygous endosperm, some loci differentially accumulate siRNAs in Pol IV heterozygous endosperm produced from Pol IV mutant mothers or fathers (Satyaki and Gehring, 2022). Maternal Pol IV mutations have a much stronger impact on endosperm siRNA accumulation and most maternally derived siRNA loci require maternal Pol IV (Satyaki and Gehring, 2022). One hypothesis to explain these observations is that RdDM establishes a heritable epigenetic state in the central cell, and this state causes siRNAs production from maternally derived alleles after fertilization (Mosher, 2010). Alternatively, maternally derived siRNAs might result from the fact that the endosperm is surrounded by, and intimately connected with, maternal sporophytic cells. As discussed above, the tapetum (male sporophytic

cells surrounding the developing male germ line) produces reproductive phasiRNA and Pol IV-dependent 24-nt siRNAs that move intercellularly into the male germ line. It is reasonable to propose that maternal sporophytic cells function similarly and continue transporting siRNAs into the endosperm throughout seed development (Figure 3C) (Grover et al., 2020). Post-fertilization movement of siRNA provides a unique opportunity for the maternal genome to respond to endosperm cues (e.g. speed of growth and development), and potentially influence gene expression to maximize embryo success. Maternal siren siRNAs have already been demonstrated to impact gene expression in *Brassica rapa* ovules (Burgess et al., 2022), although it is not known whether they also impact gene expression in endosperm, nor which gene expression changes are developmentally significant. Such a system would explain why maternal sporophytic, but not gametophytic, RdDM is required for successful seed development in *B. rapa* (Grover et al., 2018) and *Capsella rubella* (Wang et al., 2020).

Although they are hypothesized to trigger methylation in the endosperm, there is no evidence of siRNA movement from maternal soma to developing embryos. Firstly, siren siRNAs are biparental in embryos, indicating that there is not a meaningful accumulation of maternally derived siRNAs in embryos (Grover et al., 2020). Secondly, there is little difference in methylation between *rdr2* embryos whether they develop on *rdr2* mutant mothers who lack siRNA production or *rdr2/RDR2* heterozygous mother who produce somatic siRNAs (Chakraborty et al., 2021).

In addition to movement from the soma to the filial tissues, siRNAs could move between the endosperm and embryo. Just as DME-mediated demethylation of the vegetative cell causes production of siRNAs that trigger methylation in sperm cells (Ibarra et al., 2012) (discussed above), it is possible that a the demethylated endosperm provides siRNAs to the developing embryo. This hypothesis was raised based on the observed correlation between sites that are demethylated in endosperm and hypermethylated in embryos (Hsieh et al., 2009; Mosher and Melnyk, 2010; Bauer and Fischer, 2011; Bouyer et al., 2017). However, this correlation is not seen in *B. rapa* (Chakraborty et al., 2021). Furthermore, soybean somatic embryos are hypermethylated, suggesting that the signals for methylation are intrinsic to the embryogenesis program, rather than driven by demethylation of neighboring tissues (Ji et al., 2019). Although such movement is a compelling hypothesis, over a decade of research has not provided evidence for siRNA movement between endosperm and embryo, which are sympastically isolated shortly after fertilization (Lafon-Placette and Köhler, 2014). On the whole, evidence suggests that RdDM in embryos is entirely cell autonomous.

Embryo hypermethylation

During embryogenesis, the genome becomes heavily methylated at CHH sites (An et al., 2017; Bouyer et al., 2017; Kawakatsu et al., 2017; Lin et al., 2017; Narsai et al., 2017; Ji

et al., 2019; Rajkumar et al., 2020; Chakraborty et al., 2021). Indeed, mature embryos in dry seed are one of the most highly methylated plant tissues (Kawakatsu et al., 2017). Embryo hypermethylation occurs predominantly on transposons and requires RDR2 and DRM, indicating that it occurs through RdDM (Figure 3D; Lin et al., 2017; Chakraborty et al., 2021), although siRNA-independent methylation via CMT2 is also implicated in this process (Kawakatsu et al., 2017; Papareddy et al., 2020). In rice, a resetting of 24-nt siRNA production is detected in single-celled zygotes, indicating an immediate transition to an embryogenic pattern of RdDM (Li et al., 2022). Detailed analysis of siRNAs and DNA methylation throughout Arabidopsis embryogenesis indicates that heterochromatin decondensation in the embryo allows 24-nt siRNA production and subsequent CHH methylation (Papareddy et al., 2020), further supporting a model of cell-autonomous embryo methylation.

While control of transposons is important in embryos, it is not clear that embryo hypermethylation is to defend the genome against mobile elements. Firstly, the meristematic cells where transposon control is most important make up only a small fraction of cells in the mature embryo, indicating that most of the methylation is present on somatic tissues of the embryo (Papareddy et al., 2020). Secondly, methylation is lost within two days following germination, despite the fact that transposon control is still important during early growth and development (Bouyer et al., 2017; Kawakatsu et al., 2017; Narsai et al., 2017). And finally, CHH hypermethylation is dispensable for embryo development in Arabidopsis and *B. rapa* (Lin et al., 2017; Chakraborty et al., 2021). There are several theories for the function of this methylation, including maintaining a transcriptional quiescent state during dormancy (Kawakatsu et al., 2017), a side-effect of chromatin decondensation during rapid protein synthesis or cell-cycle arrest (Papareddy et al., 2020; Papareddy and Nodine, 2021). Ancient barley grains from archeological sites indicate that methylated cytosines spontaneously deaminate to thymine in dormant seeds (Smith et al., 2014), and therefore hypermethylation might be an opportunity to induce mutations in transposons while the seed awaits germination.

Trans-chromosomal methylation

Most angiosperms are outcrossing and thus the embryo results from fusion of genomes that vary both genetically and epigenetically. While gene conversion is rare, epigenetic alteration of one allele by its homologous pair is much more frequent. In its simplest form, this process is called trans-chromosomal methylation (TCM) or trans-chromosomal demethylation (TCdM) and results in DNA methylation in an F1 hybrid that diverges from the average of the two parental types (Figure 4; Greaves et al., 2012a). Loci undergoing TCM and TCdM are associated with abundant siRNAs in parental genomes, suggesting that siRNAs move between homologous chromosomes to trigger methylation at the paired allele (Chodavarapu et al., 2012; Shen et al., 2012; Greaves et al., 2012b; Zhang et al., 2016; Cao et al., 2022). Analysis of

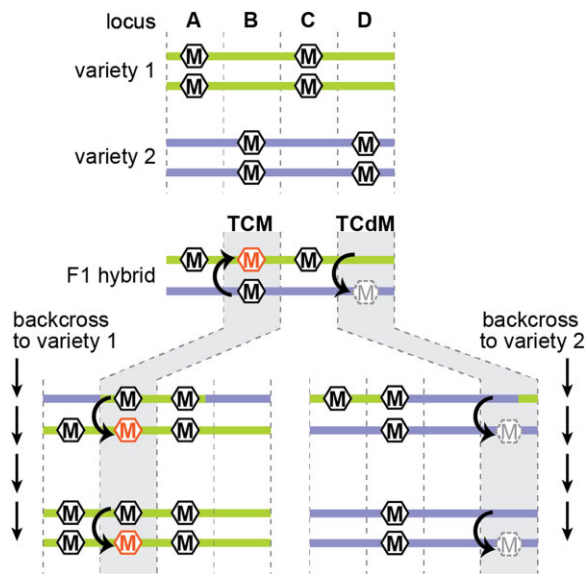


Figure 4 Trans-chromosomal methylation and demethylation. Four hypothetical loci (A–D) that are differentially methylated between two varieties. At loci A and C, there is no allelic interaction, and methylation depends on the genetic background (green is methylated, purple is unmethylated). At locus B, TCM triggers methylation on the previously unmethylated allele (orange hexagon). In repeated backcrosses to the unmethylated background, the newly methylated allele continues to induce methylation of naïve alleles. At locus D, TCdM results in demethylation of the high methylation alleles (grey dashed hexagon). This unmethylated state continues to cause TCdM in subsequent generations of backcrossing to the methylated allele.

allele-specific siRNAs demonstrates that the allele gaining methylation does not necessarily begin producing its own siRNAs (Zhang et al., 2016), further indicating that siRNAs are responsible for inter-allele communication, not just perpetuation of the newly methylated state.

On the whole, TCM is more common than TCdM, which might result when siRNAs produced from one allele divide their function between homologous alleles, resulting in a siRNA concentration that is insufficient to maintain methylation at either allele (Chodavarapu et al., 2012; Greaves et al., 2016). There is also some evidence that TCdM is more likely to occur when the alleles are more genetically dissimilar, suggesting that siRNAs might migrate to the opposite allele, but be unable to trigger methylation due to nucleotide mismatches (Zhang et al., 2016). Several recent papers demonstrate that 24-nt siRNAs can function despite mismatches between the siRNA and target site (Fei et al., 2021; Long et al., 2021; Burgess et al., 2022), but it is not clear whether mismatched siRNA operate as efficiently as perfectly matching siRNAs.

In some cases, TCM and TCdM create heritable epigenetic states, or epialleles. In crosses between two maize varieties, hundreds of TCM or TCdM events were stably maintained through six generations of backcrossing and three generations of selfing (Cao et al., 2022). Similarly, interspecific hybrids

between maize and teosinte or between tomato and its wild relative *Solanum pennellii* also induce heritable methylation changes (Shivaprasad et al., 2012; Gouil and Baulcombe, 2018; Cao et al., 2022). The stability of these changes indicates that the converted allele gains the ability to convert a naïve allele, a process called paramutation (Hollick, 2017). For example, an allele that experiences TCM becomes hypermethylated and gains the ability to transmit that high-methylation state to a low-methylation allele in the next generation (Figure 4). Heritable epialleles are associated with changes in chromatin accessibility, histone modifications, and accumulation of siRNAs (Shivaprasad et al., 2012; Hollick, 2017; Gouil and Baulcombe, 2018; Cao et al., 2022), indicating that gaining or losing the ability to produce siRNAs through changes in chromatin state might underlie the heritability some TCM and TCdM events. However, other TCM and TCdM events might be paramutagenic without the use of siRNAs (Martinho et al., 2022).

Outstanding questions

Research on small RNA-mediated DNA methylation during reproduction is an important and growing area of research. Despite many recent advances, a number of critical questions remain unanswered.

How does pol IV create tissue-specific expression of a subset of 24-nt siRNA loci?

While Pol II transcription of *PHAS* loci provides a clear mechanism for tissue-specific expression of reproductive phasiRNAs, it is unknown how expression of Pol IV-dependent siRNAs is regulated. Reproductive-specific expression of Pol IV-dependent siRNAs from some loci was first noted in 2009 and was hypothesized to result from *DME* demethylation in the maternal gametophyte (Mosher et al., 2009; Mosher, 2010). However, ectopic overexpression of *DME* was insufficient to induce siRNA expression and loss of *DME* homologs in maize is associated with increased siRNA expression (Mosher et al., 2011; Gent et al., 2022). Subsequent research suggests that chromatin decondensation might allow Pol IV access to chromatin in specific developmental states (Fu et al., 2018; Papareddy et al., 2020; Choi et al., 2021), however how decondensation might be controlled in a locus-specific manner is unclear. Both siren siRNAs in the female gametophyte and nurse cell siRNAs during male germ line development require *CLSY3* (Long et al., 2021; Zhou et al., 2022a), hinting that these chromatin remodelers, which are expressed in tissue-specific patterns, might promote site-specific decondensation to allow Pol IV access. Alternatively, decondensation and nucleosome remodeling might work together to uncover binding sites for Pol IV recruitment. Siren siRNAs in *Arabidopsis* and *B. rapa* are enriched for sequence motifs (Burgess et al., 2022; Zhou et al., 2022a), although it remains to be determined whether these motifs are functionally significant in triggering siRNA expression.

How do siRNAs move in reproductive tissues?

Although there is increasing evidence for intercellular movement of siRNAs during reproductive development, the route of this transport is unclear. siRNAs can move from cell to cell in vegetative tissue, presumably through plasmodesmata (Melnyk et al., 2011; Liu and Chen, 2018). During male reproductive development, there are many plasmodesmata connecting the tapetum to the microspore mother cell, but these are occluded during meiosis by callose deposition (Sager and Lee, 2014), suggesting that symplastic movement of 24-nt reproductive phasiRNAs or nurse cell siRNAs must conclude prior to the completion of meiosis. Similarly, plasmodesmata at connecting the megagametophyte to maternal somatic cells appear to be occluded by callose at the four nucleus stage (Thijssen, 2003) and the integuments are symplastically isolated during embryo development (Stadler et al., 2005). Lack of plasmodesmata between maternal somatic cells and the endosperm might serve to protect the endosperm and embryo from chemical intermediates released when the integuments undergo programmed cell death to create the seed coat (Sager and Lee, 2014).

Small RNAs are also found in membrane-bound extracellular vesicles (Baldrich et al., 2019; Ruf et al., 2022), providing a plausible mechanism for apoplastic movement of siRNAs. Interestingly, this pathway can transport RNA from one organism to another (Cai et al., 2018), which evokes the movement from the maternal soma to genetically distinct filial tissues. However, at this time we have only a limited understanding of intercellular small RNA transport via extracellular vesicles.

Another interesting question in the mechanism of small RNA movement is whether small RNAs move alone or in complex with RNA binding proteins. Double-stranded siRNA duplexes produced by DCL cleavage are small enough to transport through plasmodesmata, however without the protection of an RNA binding protein, “naked” siRNAs might be subject to degradation. An RNA-binding partner would also allow siRNAs to be targeted to the plasmodesmata, while unbound siRNAs would presumably move through simple diffusion. AGO proteins, the primary binding partner of siRNAs, are large proteins that are unlikely to pass through plasmodesmata (Brosnan et al., 2019; Jullien et al., 2022), but genomes contain many other RNA-binding proteins that might facilitate movement. Indeed, the specific expression of such proteins might enable tissue-specific siRNA mobility. Extracellular vesicles do not have a size exclusion limit, and several RNA-binding proteins have been identified in vesicles, including AGO1 (He et al., 2021). Discovering the mechanism of small RNA transport will undoubtedly unlock new knowledge about the nature and function of mobile small RNAs.

To what extent do reproductive siRNAs mediate heritable, transgenerational epigenetics?

Paramutagenic events following inter- or intra-specific hybridization demonstrate that epigenetic patterns in an

individual are at least partially dependent on inherited epigenetic states (Shivaprasad et al., 2012; Gouil and Baulcombe, 2018; Cao et al., 2022). And because these heritable epialleles are sometimes associated with changes in expression of neighboring genes (Cao et al., 2022), an individual's phenotype might derive from an epigenetic change induced in its ancestors. Yet most epigenetic polymorphisms do not exhibit trans-chromosomal behavior and it is unknown what properties of a locus are necessary for this chromosomal communication. It is also difficult to reconcile paramutagenic events in the face of extensive embryo hypermethylation during development and subsequent loss of methylation during germination (Bouyer et al., 2017; Kawakatsu et al., 2017; Narsai et al., 2017). Improved techniques for assaying epigenetic patterns, including single cell approaches, should enable a better understanding of how stable epigenetic patterns are transmitted.

Another outstanding question is whether siRNAs that are deposited in gametes from somatic cells influence epigenetic states in the zygote. Nurse cell siRNAs, 24-nt reproductive phasiRNAs, and easiRNAs might be loaded into the sperm cell and delivered to the egg cell upon fertilization. Alternatively, they might establish an epigenetic state on patrigenic alleles that could be transmitted to matrigenic alleles after fertilization. Similarly, it is possible that siren siRNA could accumulate in the egg cell before fertilization, or possibly be transported from maternal soma to zygote after fertilization.

How do small RNAs influence diverse plant reproductive systems?

Angiosperms are the dominant class of plants on Earth and their seeds are critically important for humans. But seed production is not the ancestral state for plants, and many extant lineages either do not make seeds, or do not make them through double fertilization as described here. It will be particularly interesting to investigate the role of small RNAs in reproductive development of bryophytes, whose lifecycle is dominated by the haploid gametophyte. For example, dynamic DNA methylation in *Marchantia polymorpha* suggests that there might be antherozoid (sperm)-specific RdDM (Schmid et al., 2018).

It will also be interesting to understand the rates of birth, death, and mutation for easiRNAs, reproductive phasiRNAs, and siren loci, which can vary even within families (Bélanger et al., 2020; Grover et al., 2020; Pokhrel et al., 2021). Understanding selection to retain these loci and maintain their ability to recognize their targets might help us understand their biological functions.

Conclusion

Advances in sequencing technology have allowed researchers to discover and catalog the many types of siRNAs produced during plant reproductive development and we are

developing an increasing understanding of how these siRNAs function. Although we have a strong grasp of how genetic information (i.e. DNA) is transmitted from parent to offspring, we still have much to learn about how epigenetic modifications are faithfully passed between generations. Epigenetic communication between generations via small RNA movement is also an exciting caveat to the standard rules of inheritance. In addition to these fundamental biological questions, understanding how small RNAs influence fertility and seed development will be critical as we attempt to increase agricultural production while minimizing environmental impacts.

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References

- An Y-QC, Goettel W, Han Q, Bartels A, Liu Z, Xiao W** (2017) Dynamic changes of genome-wide DNA methylation during soybean seed development. *Sci e* **7**(1): 12263
- Araki S, Le NT, Koizumi K, Villar-Briones A, Nonomura K-I, Endo M, Inoue H, Saze H, Komiya R** (2020) miR2118-dependent U-rich phasiRNA production in rice anther wall development. *Nat Commun* **11**(1): 3115
- Baldrich P, Rutter BD, Karimi HZ, Podicheti R, Meyers BC, Innes RW** (2019) Plant extracellular vesicles contain diverse small RNA species and are enriched in 10- to 17-nucleotide “tiny” RNAs. *Plant Cell* **31**(2): 315–324
- Bauer MJ, Fischer RL** (2011) Genome demethylation and imprinting in the endosperm. *Curr. Opin. Plant Biol* **14**(2): 162–167
- Bélanger S, Pokhrel S, Czymmek K, Meyers BC** (2020) Premeiotic, 24-nucleotide reproductive PhasiRNAs are abundant in anthers of wheat and barley but not rice and maize. *Plant Physiol* **184**(3): 1407–1423
- Berger F, Twell D** (2011) Germline specification and function in plants. *Annu Rev Plant Biol* **62**(1): 461–484
- Blevins T, Podicheti R, Mishra V, Marasco M, Wang J, Rusch D, Tang H, Pikaard CS** (2015) Identification of pol IV and RDR2-dependent precursors of 24 nt siRNAs guiding de novo DNA methylation in *Arabidopsis*. *Elife* **4**: e09591
- Böhmendorfer G, Sethuraman S, Rowley MJ, Krzyszton M, Rothi MH, Bouzit L, Wierzbicki AT** (2016) Long non-coding RNA produced by RNA polymerase V determines boundaries of heterochromatin. *Elife* **5**: e19092
- Borges F, Parent J-S, van Ex F, Wolff P, Martínez G, Köhler C, Martienssen RA** (2018) Transposon-derived small RNAs triggered by miR845 mediate genome dosage response in *Arabidopsis*. *Nat Genet* **50**(2): 186–192
- Bouyer D, Kramdi A, Kassam M, Heese M, Schnittger A, Roudier F, Colot V** (2017) DNA Methylation dynamics during early plant life. *Genome Biol* **18**(1): 179
- Brandvain Y, Haig D** (2018) Outbreeders pull harder in a parental tug-of-war. *Proc Natl Acad Sci USA* **115**(45): 11354–11356
- Brosnan CA, Sarazin A, Lim P, Bologna NG, Hirsch-Hoffmann M, Voinnet O** (2019) Genome-scale, single-cell-type resolution of microRNA activities within a whole plant organ. *EMBO J* **38**(13): e100754
- Burgess D, Chow HT, Grover JW, Freeling M, Mosher RA** (2022) Ovule siRNAs methylate protein-coding genes in trans. *Plant Cell* **34**(10): 3647–3664
- Cai Q, Qiao L, Wang M, He B, Lin F-M, Palmquist J, Huang S-D, Jin H** (2018) Plants send small RNAs in extracellular vesicles to fungal pathogen to silence virulence genes. *Science* **360**(6393): 1126–1129
- Cao S, Wang L, Han T, Ye W, Liu Y, Sun Y, Moose SP, Song Q, Chen ZJ** (2022) Small RNAs mediate transgenerational inheritance of genome-wide trans-acting epialleles in maize. *Genome Biol* **23**(1): 53
- Chakraborty T, Kendall T, Grover JW, Mosher RA** (2021) Embryo CHH hypermethylation is mediated by RdDM and is autonomously directed in *Brassica rapa*. *Genome Biol* **22**(1): 140
- Chakraborty T, Trujillo JT, Kendall T, Mosher RA** (2022) A null allele of the pol IV second subunit impacts stature and reproductive development in *Oryza sativa*. *Plant J* **111**(3): 748–755
- Chan SW-L, Zhang X, Bernatavichute YV, Jacobsen SE** (2006) Two-step recruitment of RNA-directed DNA methylation to tandem repeats. *PLoS Biol* **4**(11): e363
- Chodavarapu RK, Feng S, Ding B, Simon SA, Lopez D, Jia Y, Wang G-L, Meyers BC, Jacobsen SE, Pellegrini M** (2012) Transcriptome and methylome interactions in rice hybrids. *Proc Natl Acad Sci USA* **109**(30): 12040–12045
- Choi J, Lyons DB, Zilberman D** (2021) Histone H1 prevents non-CG methylation-mediated small RNA biogenesis in *Arabidopsis* heterochromatin. *Elife* **10**: e72676
- Creasey KM, Zhai J, Borges F, Van Ex F, Regulski M, Meyers BC, Martienssen RA** (2014) miRNAs trigger widespread epigenetically activated siRNAs from transposons in *Arabidopsis*. *Nature* **508** (7496): 411–415
- Cuerda-Gil D, Slotkin RK** (2016) Non-canonical RNA-directed DNA methylation. *Nat Plants* **2**(11): 16163
- Erdmann RM, Satyaki PRV, Klosinska M, Gehring M** (2017) A small RNA pathway mediates allelic dosage in endosperm. *Cell Rep* **21**(12): 3364–3372
- Erhard, K.F., Jr, Stonaker, J.L., Parkinson, S.E., Lim, J.P., Hale, C.J., and Hollick, J.B.** (2009). RNA Polymerase IV functions in paramutation in *Zea mays*. *Science* **323**(5918): 1201–1205
- Fei Y, Nyikó T, Molnar A** (2021) Non-perfectly matching small RNAs can induce stable and heritable epigenetic modifications and can be used as molecular markers to trace the origin and fate of silencing RNAs. *Nucleic Acids Res* **49**(4): 1900–1913
- Feng X, Zilberman D, Dickinson H** (2013) A conversation across generations: soma-germ cell crosstalk in plants. *Dev Cell* **24**(3): 215–225
- Fu F-F, Dawe RK, Gent JI** (2018) Loss of RNA-directed DNA methylation in maize chromomethylase and DDM1-type nucleosome remodeler mutants. *Plant Cell* **30**(7): 1617–1627
- Fukudome A, Singh J, Mishra V, Reddem E, Martínez-Marquez F, Wenzel S, Yan R, Shiozaki M, Yu Z, Wang J-C-Y, et al.** (2021) Structure and RNA template requirements of *Arabidopsis* RNA-DEPENDENT RNA POLYMERASE 2. *Proc Natl Acad Sci* **118**(51): e2115899118
- Gent JI, Higgins KM, Swentowsky KW, Fu F-F, Zeng Y, Kim DW, Dawe RK, Springer NM, Anderson SN** (2022) The maize gene maternal derepression of r1 encodes a DNA glycosylase that demethylates DNA and reduces siRNA expression in the endosperm. *Plant Cell* **34**(10): 3685–3701
- Gouil Q, Baulcombe DC** (2016) DNA Methylation signatures of the plant chromomethyltransferases. *PLoS Genet* **12**(12): e1006526
- Gouil Q, Baulcombe DC** (2018) Paramutation-like features of multiple natural epialleles in tomato. *BMC Genom* **19**(1):203
- Greaves IK, Eichten SR, Groszmann M, Wang A, Ying H, Peacock WJ, Dennis ES** (2016) Twenty-four-nucleotide siRNAs produce heritable trans-chromosomal methylation in F1 *Arabidopsis* hybrids. *Proc Natl Acad Sci USA* **113**(44): E6895–E6902
- Greaves I, Groszmann M, Dennis ES, Peacock WJ** (2012a) Trans-chromosomal methylation. *Epigenetics* **7**(8): 800–805
- Greaves IK, Groszmann M, Ying H, Taylor JM, Peacock WJ, Dennis ES** (2012b) Trans chromosomal methylation in *Arabidopsis* hybrids. *Proc Natl Acad Sci USA* **109**(9): 3570–3575

- Grover JW, Burgess D, Kendall T, Baten A, Pokhrel S, King GJ, Meyers BC, Freeling M, Mosher RA (2020) Abundant expression of maternal siRNAs is a conserved feature of seed development. *Proc Natl Acad Sci USA* **117**(26): 15305–15315
- Grover JW, Kendall T, Baten A, Burgess D, Freeling M, King GJ, Mosher RA (2018) Maternal components of RNA-directed DNA methylation are required for seed development in *Brassica rapa*. *Plant J* **94**(4): 575–582
- Haig D, Westoby M (1989) Parent-specific gene expression and the triploid endosperm. *Am Nat* **134**(1): 147–155
- He B, Cai Q, Qiao L, Huang C-Y, Wang S, Miao W, Ha T, Wang Y, Jin H (2021). RNA-binding proteins contribute to small RNA loading in plant extracellular vesicles. *Nat Plants* **7**(3): 342
- Henderson IR, Jacobsen SE (2008) Tandem repeats upstream of the Arabidopsis endogene SDC recruit non-CG DNA methylation and initiate siRNA spreading. *Genes Dev* **22**(12): 1597–1606
- Hollick JB (2017) Paramutation and related phenomena in diverse species. *Nat Rev Genet* **18**(1): 5–23
- Hollister JD, Gaut BS (2009) Epigenetic silencing of transposable elements: a trade-off between reduced transposition and deleterious effects on neighboring gene expression. *Genome Res* **19**(8): 1419–1428
- Hsieh T-F, Ibarra CA, Silva P, Zemach A, Eshed-Williams L, Fischer RL, Zilberman D (2009) Genome-wide demethylation of Arabidopsis endosperm. *Science* **324**(5933): 1451–1454
- Huang K, Wu X-X, Fang C-L, Xu Z-G, Zhang H-W, Gao J, Zhou C-M, You L-L, Gu Z-X, Mu W-H, et al. (2021) Pol IV and RDR2: a two-RNA-polymerase machine that produces double-stranded RNA. *Science* **374**(6575): 1579–1586
- Ibarra CA, Feng X, Schoft VK, Hsieh TF, Uzawa R, Rodrigues JA, Zemach A, Chumak N, Machlicova A, Nishimura T, Rojas D, et al. (2012) Active DNA demethylation in plant companion cells reinforces transposon methylation in gametes. *Science* **337**(6100): 1360–1364
- Ingouff M, Selles B, Michaud C, Vu TM, Berger F, Schorn AJ, Autran D, Van Durme M, Nowack MK, Martienssen RA, et al. (2017) Live-cell analysis of DNA methylation during sexual reproduction in Arabidopsis reveals context and sex-specific dynamics controlled by noncanonical RdDM. *Genes Dev* **31**(1): 72–83
- Ji L, Mathioni SM, Johnson S, Tucker D, Bewick AJ, Do Kim K, Daron J, Slotkin RK, Jackson SA, Parrott WA, et al. (2019) Genome-wide reinforcement of DNA methylation occurs during somatic embryogenesis in soybean. *Plant Cell* **31**(10): 2315–2331
- Jiang P, Lian B, Liu C, Fu Z, Shen Y, Cheng Z, Qi Y (2020) 21-nt phasiRNAs direct target mRNA cleavage in rice male germ cells. *Nat Commun* **11**(1): 5191
- Johnson C, Kasprzewska A, Tennesen K, Fernandes J, Nan G-L, Walbot V, Sundaresan V, Vance V, Bowman LH (2009) Clusters and superclusters of phased small RNAs in the developing inflorescence of rice. *Genome Res* **19**(8): 1429–1440
- Johnston, S.A., den Nijs TP, Peloquin SJ, Hanneman RE Jr (1980). The significance of genic balance to endosperm development in interspecific crosses. *Theor Appl Genet* **57**(1): 5–9
- Jullien PE, Schröder JA, Bonnet DMV, Pumplun N, Voinnet O (2022) Asymmetric expression of argonautes in reproductive tissues. *Plant Physiol* **188**(1): 38–43
- Kakrana A, Mathioni SM, Huang K, Hammond R, Vandivier L, Patel P, Arikis S, Shevchenko O, Harkess AE, Kingham B, et al. (2018) Plant 24-nt reproductive phasiRNAs from intramolecular duplex mRNAs in diverse monocots. *Genome Res* **28**(9): 1333–1344
- Kasschau KD, Fahlgren N, Chapman EJ, Sullivan CM, Cumbie JS, Givan SA, Carrington JC (2007) Genome-wide profiling and analysis of Arabidopsis siRNAs. *PLoS Biol* **5**(3): e57
- Kawakatsu T, Nery JR, Castanon R, Ecker JR (2017) Dynamic DNA methylation reconfiguration during seed development and germination. *Genome Biol* **18**(1): 171
- Khouider S, Borges F, LeBlanc C, Ungru A, Schnitger A, Martienssen R, Colot V, Bouyer D (2021) Male fertility in Arabidopsis requires active DNA demethylation of genes that control pollen tube function. *Nat Commun* **12**(1): 410
- Kim MY, Ono A, Scholten S, Kinoshita T, Zilberman D, Okamoto T, Fischer RL (2019) DNA demethylation by ROS1a in rice vegetative cells promotes methylation in sperm. *Proc Natl Acad Sci USA* **116**(19): 9652–9657
- Kinoshita Y, Saze H, Kinoshita T, Miura A, Soppe WJJ, Koornneef M, Kakutani T (2007) Control of FWA gene silencing in Arabidopsis thaliana by SINE-related direct repeats. *Plant J* **49**(1): 38–45
- Kirkbride RC, Lu J, Zhang C, Mosher RA, Baulcombe DC, Chen ZJ (2019) Maternal small RNAs mediate spatial-temporal regulation of gene expression, imprinting, and seed development in Arabidopsis. *Proc Natl Acad Sci USA* **116**(7): 2761–2766
- Kradolfer D, Wolff P, Jiang H, Siretskiy A, Köhler C (2013) An imprinted gene underlies postzygotic reproductive isolation in Arabidopsis thaliana. *Dev Cell* **26**(5): 525–535
- Lafon-Placette C, Köhler C (2014) Embryo and endosperm, partners in seed development. *Curr Opin Plant Biol* **17**: 64–69
- Lahmy S, Pontier D, Bies-Etheve N, Laudie M, Feng S, Jobet E, Hale CJ, Cooke R, Hakimi M-A, Angelov D, et al. (2016) Evidence for ARGONAUTE4-DNA interactions in RNA-directed DNA methylation in plants. *Genes Dev* **30**(23): 2565–2570
- Law JA, Du J, Hale CJ, Feng S, Krajewski K, Palanca AMS, Strahl BD, Patel DJ, Jacobsen SE (2013) Polymerase IV occupancy at RNA-directed DNA methylation sites requires SHH1. *Nature* **498**(7454): 385–389
- Law JA, Jacobsen SE (2010) Establishing, maintaining and modifying DNA methylation patterns in plants and animals. *Nat Rev Genet* **11**(3): 204–220
- Li Q, et al. (2015) RNA-directed DNA methylation enforces boundaries between heterochromatin and euchromatin in the maize genome. *Proc Natl Acad Sci USA* **112**(47): 14728–14733
- Li C, Gent JI, Xu H, Fu H, Russell SD, Sundaresan V (2022) Resetting of the 24-nt siRNA landscape in rice zygotes. *Genome Res* **32**(2): 309–323
- Li C, Xu H, Fu F-F, Russell SD, Sundaresan V, Gent JI (2020) Genome-wide redistribution of 24-nt siRNAs in rice gametes. *Genome Res* **30**(2): 173–184
- Lin J-Y, Le BH, Chen M, Henry KF, Hur J, Hsieh T-F, Chen P-Y, Pelletier JM, Pellegrini M, Fischer RL, et al. (2017) Similarity between soybean and Arabidopsis seed methylomes and loss of non-CG methylation does not affect seed development. *Proc Natl Acad Sci USA* **114**(45): E9730–E9739
- Liu L, Chen X (2018) Intercellular and systemic trafficking of RNAs in plants. *Nat Plants* **4**(11): 869–878
- Liu W, Duttke SH, Hetzel J, Groth M, Feng S, Gallego-Bartolome J, Zhong Z, Kuo HY, Wang Z, Zhai J, et al. (2018) RNA-directed DNA methylation involves co-transcriptional small-RNA-guided slicing of polymerase V transcripts in Arabidopsis. *Nat Plants* **4**(3): 181–188
- Loffer A, Singh J, Fukudome A, Mishra V, Wang F, Pikaard CS (2022) A DCL3 dicing code within pol IV-RDR2 transcripts diversifies the siRNA pool guiding RNA-directed DNA methylation. *Elife* **11**: e73260
- Long J, Walker J, She W, Aldridge B, Gao H, Deans S, Vickers M, Feng X (2021) Nurse cell-derived small RNAs define paternal epigenetic inheritance in Arabidopsis. *Science* **373**(6550): eabh0556
- Lu J, Zhang C, Baulcombe DC, Chen ZJ (2012) Maternal siRNAs as regulators of parental genome imbalance and gene expression in endosperm of Arabidopsis seeds. *Proc Natl Acad Sci USA* **109**(14): 5529–5534
- Martinez G, Wolff P, Wang Z, Moreno-Romero J, Santos-González J, Conze LL, DeFraia C, Slotkin RK, Köhler C (2018) Paternal easiRNAs regulate parental genome dosage in Arabidopsis. *Nat Genet* **50**(2): 193–198
- Martinho C, Wang Z, Ghigi A, Buddle S, Barbour F, Yarur A, Gouil Q, Müller S, Stam M, Liu C, et al. (2022) CHROMOMETHYLTRANSFERASE3/KRYPTONITE Maintains the

- sulfurea paramutation in *Solanum lycopersicum*. *Proc Natl Acad Sci USA* **119**(13): e2112240119
- Martínez G, Panda K, Köhler C, Slotkin RK** (2016) Silencing in sperm cells is directed by RNA movement from the surrounding nurse cell. *Nat Plants* **2**(4): 16030
- Matzke MA, Mosher RA** (2014) RNA-directed DNA methylation: an epigenetic pathway of increasing complexity. *Nat. Rev Genet* **15**(6): 394–408
- McCue AD, Panda K, Nuthikattu S, Choudury SG, Thomas EN, Slotkin RK** (2015) ARGONAUTE 6 Bridges transposable element mRNA-derived siRNAs to the establishment of DNA methylation. *EMBO J* **34**(1): 20–35
- Melnyk CW, Molnar A, Baulcombe DC** (2011) Intercellular and systemic movement of RNA silencing signals. *EMBO J* **30**(17): 3553–3563
- Mendes MA, Petrella R, Cucinotta M, Vignati E, Gatti S, Pinto SC, Bird DC, Gregis V, Dickinson H, Tucker MR, et al.** (2020) The RNA-dependent DNA methylation pathway is required to restrict SPOROCTELESS/NOZZLE expression to specify a single female germ cell precursor in *Arabidopsis*. *Development* **147**(23): dev194274
- Mosher RA** (2010) Maternal control of Pol IV-dependent siRNAs in *Arabidopsis* endosperm. *New Phytol* **186**(2): 358–364
- Mosher RA, Melnyk CW** (2010) siRNAs and DNA methylation: seedy epigenetics. *Trends Plant Sci* **15**(4): 204–210
- Mosher RA, Melnyk CW, Kelly KA, Dunn RM, Studholme DJ, Baulcombe DC** (2009) Uniparental expression of PolIV-dependent siRNAs in developing endosperm of *Arabidopsis*. *Nature* **460**(7252): 283–286
- Mosher RA, Schwach F, Studholme D, Baulcombe DC** (2008) PolIVb influences RNA-directed DNA methylation independently of its role in siRNA biogenesis. *Proc Natl Acad Sci USA* **105**(8): 3145–3150
- Mosher RA, Tan EH, Shin J, Fischer RL, Pikaard CS, Baulcombe DC** (2011) An atypical epigenetic mechanism affects uniparental expression of Pol IV-dependent siRNAs. *PLoS One* **6**(10): e25756
- Müntzing A** (1933) Hybrid incompatibility and the origin of polyploidy. *Hereditas* **18**(1–2): 33–55
- Nan G-L, Teng C, Fernandes J, O'Connor L, Meyers BC, Walbot V** (2022) A cascade of bHLH-regulated pathways programs maize anther development. *Plant Cell* **34**(4): 1207–1225
- Narsai R, Secco D, Schultz MD, Ecker JR, Lister R, Whelan J** (2017) Dynamic and rapid changes in the transcriptome and epigenome during germination and in developing rice (*Oryza sativa*) coleoptiles under anoxia and re-oxygenation. *Plant J* **89**(4): 805–824
- Nuthikattu S, McCue AD, Panda K, Fultz D, DeFraia C, Thomas EN, Slotkin RK** (2013) The initiation of epigenetic silencing of active transposable elements is triggered by RDR6 and 21–22 nucleotide small interfering RNAs. *Plant Physiol* **162**(1): 116–131
- Olmedo-Monfil V, Durán-Figueroa N, Arteaga-Vázquez M, Demesa-Arévalo E, Autran D, Grimanelli D, Slotkin RK, Martienssen RA, Vielle-Calzada J-P** (2010) Control of female gamete formation by a small RNA pathway in *Arabidopsis*. *Nature* **464**(7288): 628–632
- Panda K, McCue AD, Slotkin RK** (2020) *Arabidopsis* RNA polymerase IV generates 21–22 nucleotide small RNAs that can participate in RNA-directed DNA methylation and may regulate genes. *Philos Trans R Soc Lond B Biol Sci* **375**(1795): 20190417
- Papareddy RK, Nodine MD** (2021) Plant epigenetics: propelling DNA methylation variation across the cell cycle. *Curr Biol* **31**(3): R129–R131
- Papareddy RK, Páldi K, Paulraj S, Kao P, Lutzmayer S, Nodine MD** (2020) Chromatin regulates expression of small RNAs to help maintain transposon methylome homeostasis in *Arabidopsis*. *Genome Biol* **21**(1): 251
- Park J-S, Frost JM, Park K, Ohr H, Park GT, Kim S, Eom H, Lee I, Brooks JS, Fischer RL, et al.** (2017) Control of DEMETER DNA demethylase gene transcription in male and female gamete companion cells in *Arabidopsis thaliana*. *Proc Natl Acad Sci USA* **114**(8): 2078–2083
- Pignatta D, Erdmann RM, Scheer E, Picard CL, Bell GW, Gehring M** (2015) Correction: natural epigenetic polymorphisms lead to intraspecific variation in *Arabidopsis* gene imprinting. *Elife* **4**: e08658
- Pokhrel S, Huang K, Bélanger S, Zhan J, Caplan JL, Kramer EM, Meyers BC** (2021) Pre-meiotic 21-nucleotide reproductive phasiRNAs emerged in seed plants and diversified in flowering plants. *Nat Commun* **12**(1): 4941
- Pokhrel S, Meyers BC** (2022) Heat-responsive microRNAs and phased small interfering RNAs in reproductive development of flax. *Plant Direct* **6**(2): e385
- Queller DC, Strassmann JE** (2002) The many selves of social insects. *Science* **296**(5566): 311–313
- Rajkumar MS, Gupta K, Khemka NK, Garg R, Jain M** (2020) DNA methylation reprogramming during seed development and its functional relevance in seed size/weight determination in chickpea. *Commun Biol* **3**(1): 340
- Raunsgard A, Opedal ØH, Ekrem RK, Wright J, Bolstad GH, Armbruster WS, Pélabon C** (2018) Intersexual conflict over seed size is stronger in more outcrossed populations of a mixed-mating plant. *Proc Natl Acad Sci USA* **115**(45): 11561–11566
- Rodrigues JA, Ruan R, Nishimura T, Sharma MK, Sharma R, Ronald PC, Fischer RL, Zilberman D** (2013) Imprinted expression of genes and small RNA is associated with localized hypomethylation of the maternal genome in rice endosperm. *Proc Natl Acad Sci USA* **110**(19): 7934–7939
- Ruf A, Oberkofler L, Robatzek S, Weiberg A** (2022) Spotlight on plant RNA-containing extracellular vesicles. *Curr Opin Plant Biol* **69**: 102272
- Sager R, Lee J-Y** (2014) Plasmodesmata in integrated cell signalling: insights from development and environmental signals and stresses. *J Exp Bot* **65**(22): 6337–6358
- Satyaki PRV, Gehring M** (2019) Paternally acting canonical RNA-directed DNA methylation pathway genes sensitize *Arabidopsis* endosperm to paternal genome dosage. *Plant Cell* **31**(7): 1563–1578
- Satyaki PRV, Gehring M** (2022) RNA Pol IV induces antagonistic parent-of-origin effects on *Arabidopsis* endosperm. *PLoS Biol* **20**(4): e3001602
- Schmid MW, Giraldo-Fonseca A, Rövekamp M, Smetanin D, Bowman JL, Grossniklaus U** (2018) Extensive epigenetic reprogramming during the life cycle of *Marchantia polymorpha*. *Genome Biol* **19**(1): 9
- Schoft VK, Chumak N, Choi Y, Hannon M, Garcia-Aguilar M, Machlicova A, Slusarz L, Mosiolek M, Park J-S, Park GT, et al.** (2011) Function of the DEMETER DNA glycosylase in the *Arabidopsis thaliana* male gametophyte. *Proc Natl Acad Sci USA* **108**(19): 8042–8047
- Schon MA, Nodine MD** (2017) Widespread contamination of *Arabidopsis* embryo and endosperm transcriptome data sets. *Plant Cell* **29**(4): 608–617
- Scott RJ, Spielman M, Bailey J, Dickinson HG** (1998) Parent-of-origin effects on seed development in *Arabidopsis thaliana*. *Development* **125**(17): 3329–3341
- Shen H, He H, Li J, Chen W, Wang X, Guo L, Peng Z, He G, Zhong S, Qi Y, et al.** (2012) Genome-wide analysis of DNA methylation and gene expression changes in two *Arabidopsis* ecotypes and their reciprocal hybrids. *Plant Cell* **24**(3): 875–892
- Shivaprasad PV, Dunn RM, Santos BA, Bassett A, Baulcombe DC** (2012) Extraordinary transgressive phenotypes of hybrid tomato are influenced by epigenetics and small silencing RNAs. *EMBO J* **31**(2): 257–266
- Sigman MJ, Panda K, Kirchner R, McLain LL, Payne H, Peasari JR, Husbands AY, Slotkin RK, McCue AD** (2021) An siRNA-guided argonaute protein directs RNA polymerase V to initiate DNA methylation. *Nat Plants* **7**(11): 1461–1474
- Singh J, Mishra V, Wang F, Huang H-Y, Pikaard CS** (2019) Reaction mechanisms of pol IV, RDR2, and DCL3 drive RNA channeling in

- the siRNA-directed DNA methylation pathway. *Mol Cell* **75**(3): 576–589.e5
- Slotkin RK, Vaughn M, Borges F, Tanurdzić M, Becker JD, Feijó JA, Martienssen RA** (2009) Epigenetic reprogramming and small RNA silencing of transposable elements in pollen. *Cell* **136**(3): 461–472
- Smith O, Clapham AJ, Rose P, Liu Y, Wang J, Allaby RG** (2014) Genomic methylation patterns in archaeological barley show de-methylation as a time-dependent diagenetic process. *Sci Rep* **4**(1): 5559
- Song X, et al.** (2012) Roles of DCL4 and DCL3b in rice phased small RNA biogenesis. *Plant J* **69**(3): 462–474
- Stadler R, Lauterbach C, Sauer N** (2005) Cell-to-cell movement of green fluorescent protein reveals post-phloem transport in the outer integument and identifies symplastic domains in Arabidopsis seeds and embryos. *Plant Physiol* **139**(2): 701–712
- Stroud H, Do T, Du J, Zhong X, Feng S, Johnson L, Patel DJ, Jacobsen SE** (2014) Non-CG methylation patterns shape the epigenetic landscape in Arabidopsis. *Nat Struct Mol Biol* **21**(1): 64–72
- Su Z, Wang N, Hou Z, Li B, Li D, Liu Y, Cai H, Qin Y, Chen X** (2020) Regulation of female germline specification via small RNA mobility in Arabidopsis. *Plant Cell* **32**(9): 2842–2854
- Su Z, Zhao L, Zhao Y, Li S, Won S, Cai H, Wang L, Li Z, Chen P, Qin Y, et al.** (2017) The THO complex non-cell-autonomously represses female germline specification through the TAS3-ARF3 module. *Curr Biol* **27**(11): 1597–1609.e2
- Teng C, Zhang H, Hammond R, Huang K, Meyers BC, Walbot V** (2020) Dicer-like 5 deficiency confers temperature-sensitive male sterility in maize. *Nat Commun* **11**(1): 2912
- Thijssen MH** (2003) Ovules, megagametophytes and embryos. Ultrastructural studies after cryofixation. Wageningen University and Research
- Vu TM, Nakamura M, Calarco JP, Susaki D, Lim PQ, Kinoshita T, Higashiyama T, Martienssen RA, Berger F** (2013) RNA-directed DNA methylation regulates parental genomic imprinting at several loci in Arabidopsis. *Development* **140**(14): 2953–2960
- Walker J, Gao H, Zhang J, Aldridge B, Vickers M, Higgins JD, Feng X** (2018) Sexual-lineage-specific DNA methylation regulates meiosis in Arabidopsis. *Nat Genet* **50**(1): 130–137
- Wang Q, et al.** (2021) Mechanism of siRNA production by a plant dicer-RNA complex in dicing-competent conformation. *Science* **374**(6571): 1152–1157
- Wang Z, Butel N, Santos-González J, Borges F, Yi J, Martienssen RA, Martinez G, Köhler C** (2020) Polymerase IV plays a crucial role in pollen development in *Capsella*. *Plant Cell* **32**(4): 950–966
- Wang L, Zheng K, Zeng L, Xu D, Zhu T, Yin Y, Zhan H, Wu Y, Yang D-L** (2022) Reinforcement of CHH methylation through RNA-directed DNA methylation ensures sexual reproduction in rice. *Plant Physiol* **188**(2): 1189–1209
- Wierzbicki AT, Haag JR, Pikaard CS** (2008) Noncoding transcription by RNA polymerase pol IVb/pol V mediates transcriptional silencing of overlapping and adjacent genes. *Cell* **135**(4): 635–648
- Wierzbicki AT, Ream TS, Haag JR, Pikaard CS** (2009) RNA Polymerase V transcription guides ARGONAUTE4 to chromatin. *Nat Genet* **41**(5): 630–634
- Wolff P, Jiang H, Wang G, Santos-González J, Köhler C** (2015) Paternally expressed imprinted genes establish postzygotic hybridization barriers in Arabidopsis thaliana. *Elife* **4**: e10074
- Xia R, Chen C, Pokhrel S, Ma W, Huang K, Patel P, Wang F, Xu J, Liu Z, Li J, et al.** (2019) 24-nt Reproductive phasiRNAs are broadly present in angiosperms. *Nat Commun* **10**(1): 627
- Xin M, Yang R, Yao Y, Ma C, Peng H, Sun Q, Wang X, Ni Z** (2014) Dynamic parent-of-origin effects on small interfering RNA expression in the developing maize endosperm. *BMC Plant Biol* **14**(1): 192
- Xu L, Yuan K, Yuan M, Meng X, Chen M, Wu J, Li J, Qi Y** (2020) Regulation of rice tillering by RNA-directed DNA methylation at miniature inverted-repeat transposable elements. *Mol Plant* **13**(6): 851–863
- Yadava P, Tamim S, Zhang H, Teng C, Zhou X, Meyers BC, Walbot V** (2021) Transgenerational conditioned male fertility of HD-ZIP IV transcription factor mutant ocl4: impact on 21-nt phasiRNA accumulation in pre-meiotic maize anthers. *Plant Reprod* **34**(2): 117–129
- Yu L, Guo R, Jiang Y, Ye X, Yang Z, Meng Y, Shao C** (2021) Degradome sequencing-based identification of phasiRNAs biogenesis pathways in *Oryza sativa*. *BMC Genomics* **22**(1): 93
- Zemach A, Kim MY, Hsieh P-H, Coleman-Derr D, Eshed-Williams L, Thao K, Harmer SL, Zilberman D** (2013) The Arabidopsis nucleosome remodeler DDM1 allows DNA methyltransferases to access H1-containing heterochromatin. *Cell* **153**(1): 193–205
- Zhai J, Bischof S, Wang H, Feng S, Lee T-F, Teng C, Chen X, Park SY, Liu L, Gallego-Bartolome J, et al.** (2015a) A one precursor one siRNA model for pol IV-dependent siRNA biogenesis. *Cell* **163**(2): 445–455
- Zhai J, Zhang H, Arikat S, Huang K, Nan G-L, Walbot V, Meyers BC** (2015b) Spatiotemporally dynamic, cell-type-dependent premeiotic and meiotic phasiRNAs in maize anthers. *Proc Natl Acad Sci USA* **112**(10): 3146–3151
- Zhang X, Henderson IR, Lu C, Green PJ, Jacobsen SE** (2007) Role of RNA polymerase IV in plant small RNA metabolism. *Proc Natl Acad Sci USA* **104**: 4536–4541
- Zhang M, Ma X, Wang C, Li Q, Meyers BC, Springer NM, Walbot V** (2021) CHH DNA methylation increases at 24-PHAS loci depend on 24-nt phased small interfering RNAs in maize meiotic anthers. *New Phytol* **229**: 2984–2997
- Zhang Q, Wang D, Lang Z, He L, Yang L, Zeng L, Li Y, Zhao C, Huang H, Zhang H, et al.** (2016) Methylation interactions in Arabidopsis hybrids require RNA-directed DNA methylation and are influenced by genetic variation. *Proc Natl Acad Sci USA* **113**(29): E4248–E4256
- Zheng K, Wang L, Zeng L, Xu D, Guo Z, Gao X, Yang D-L** (2021) The effect of RNA polymerase V on 24-nt siRNA accumulation depends on DNA methylation contexts and histone modifications in rice. *Proc Natl Acad Sci USA* **118**(30): e2100709118
- Zhong X, Du J, Hale CJ, Gallego-Bartolome J, Feng S, Vashisht AA, Chory J, Wohlschlegel JA, Patel DJ, Jacobsen SE** (2014) Molecular mechanism of action of plant DRM de novo DNA methyltransferases. *Cell* **157**: 1050–1060
- Zhou M, Coruh C, Xu G, Martins LM, Bourbousse C, Lamboloz A, Law JA** (2022a) The CLASSY family controls tissue-specific DNA methylation patterns in Arabidopsis. *Nat Commun* **13**(1): 244
- Zhou X, Huang K, Teng C, Abdelgawad A, Batish M, Meyers BC, Walbot V** (2022b) 24-nt phasiRNAs move from tapetal to meiotic cells in maize anthers. *New Phytol* **235**(2): 488–501
- Zhou M, Palanca AMS, Law JA** (2018) Locus-specific control of the de novo DNA methylation pathway in Arabidopsis by the CLASSY family. *Nat Genet* **50**(6): 865–873